

Design, Fabrication and Characterization of Transdermal Patches Using Different Natural Polymers Containing Caffeine and Ibuprofen for Long-Term Management of Migraine

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ABSTRACT

Background: The study explores transdermal drug delivery as a solution to traditional oral and parenteral administration challenges, highlighting improvements in efficiency and scope with micro needle technology. **Objectives:** The primary objective of this study was to fabricate a natural gum-grafted matrix transdermal patch for prolonged migraine treatment, emphasizing its *in vitro* characterization. **Materials and Methods:** The materials utilized included active pharmaceutical ingredients (Ibuprofen, Caffeine), polymers (HPMC, Tamarind and Moringa), solvent (distilled water), permeation enhancer (Tween-80 5%w/w) and a backing layer. For the fabrication of the patches modified solvent evaporation method was employed. **Results and Discussion:** The physical characteristics, including folding endurance and drug content, are within the official limit. The thermal analysis shows its excellent thermal stability. The *in vitro* drug release shows the maximum 98.3% drug release up to 24 hr, while the release kinetic shows its sustained behavior upon applying different kinetic models. **Conclusion:** In conclusion, this innovative and economical transdermal approach holds promise for prolonged migraine management for up to 24 hr, offering a potential alternative to address challenges associated with traditional delivery methods used.

Keywords: Migraine, Ibuprofen, Transdermal Patch, Fabrication, Sustained release matrix system.

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INTRODUCTION

The skin, the body's largest organ, acts as a drug administration route and protective barrier against external threats, comprising the epidermis, dermis and hypodermis.¹ Transdermal Drug Delivery (TDD) is a painless, effective method for administering the drug through intact skin, ensuring a consistent pharmacokinetic profile, reducing peak levels and minimizing toxic side effects; especially beneficial for unconscious, vomiting or self-administering patients.²

A Transdermal Drug Delivery System (TDDS) is a medical device that administers medication directly into the bloodstream, ensuring controlled and extended-release. It minimizes gastrointestinal side effects and offers a convenient, non-invasive

method. Advancements in this field reflect safe and efficient drug administration.^{2,3,4}

Permeation enhancers are used in transdermal patch formulations to improve drug absorption by temporarily altering the skin barrier. They enhance drug solubility, pH and diffusion by increasing skin permeability, enhancing drug solubility and enhancing drug absorption.^{2,5,6}

Understanding skin permeation kinetics is crucial for developing transdermal patches, which affect drug permeation rate; lag time and steady-state flux. Techniques like Franz diffusion cells and tape stripping can be used to study these parameters.^{2,5,7}

A patch typically consists of a laminate structure with a backing layer, adhesive layer and drug reservoir layer, providing support and protection. Transdermal patches are a convenient and effective alternative to oral administration for certain drugs, such as insulin, methotrexate, lidocaine/prilocaine, caffeine, pilocarpine, apomorphine, paroxetine, iodine, verapamil and sumatriptan. However, they have limitations such as inadequate diffusion of large molecules, potential skin irritation, higher cost,



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limited permeability for hydrophilic drugs, discomfort during use, molecular weight constraints and unsuitability for all skin types. The development of transdermal patches for these drugs requires careful consideration of factors such as physicochemical properties, skin permeation characteristics and therapeutic efficacy.^{8,9}

Migraines can be treated with various medications, including analgesics like Ibuprofen and caffeine. Ibuprofen is a selective inhibitor of COX-1 and COX-2, while caffeine enhances cognitive performance and exercise endurance. Matrix-forming polymers like HPMC, Ethylcellulose and Sodium CMC can be used. Other drugs, such as insulin, methotrexate, lidocaine/prilocaine and caffeine, can be formulated for transdermal delivery, providing a convenient and effective alternative to other forms of administration. However, careful consideration of physicochemical properties, skin permeation characteristics and therapeutic efficacy is required.¹⁰ The aims and objectives behind our research were to fabricate a Moringa and Tamarind gum grafted modified reservoir-matrix transdermal patch for prolonged management of Migraine and its *in vitro* characterization.

MATERIALS AND METHODS

Materials

In the fabrication of transdermal patches, two natural polymers, Moringa and Tamarind gum, were employed alongside a semi-synthetic or modified polymer, Hydroxypropyl Methyl Cellulose (HPMC), Distilled water, procured from the Sigma Aldrich Germany distillation apparatus, served as the solvent. The solvent selection was a critical consideration, focusing on its ability to efficiently dissolve both the polymer and the drug, coupled with its propensity to evaporate completely without leaving any residue. Caffeine and Ibuprofen were selected as therapeutic agents for migraine management. Sodium Lauryl Sulfate (SLS), acting as a surfactant and permeation enhancer. The inclusion of Tween-80 (Polysorbate 80) was aimed to enhance permeation and functions as a non-ionic surfactant and emulsifier, particularly beneficial for improving Ibuprofen solubility. Lastly, Sodium Benzoate served as a preservative, ensuring stability throughout their shelf-life. All materials were procured from Sigma-Aldrich, meeting pharmaceutical-grade standards.¹¹

Preformulation studies

Physicochemical characterization of drugs

These studies focus on the physicochemical studies of the drug substance that may affect drug performance and development. They also provide essential information for formulation design and molecular modification. Physicochemical drug characterization was done as the melting point was determined

using MP apparatus by taking a small sample of the drug in a sealed capillary and observing it under light using a thermometer, solubility of the selected drug was determined in distilled water at room temperature, partition coefficient was determined using n-octanol as oil phase and Phosphate buffer solution (pH 7.4) as aqueous phase, calibration curves were prepared using the wavelength of 273 nm and 222 nm for Caffeine and Ibuprofen respectively, The stability studies were conducted by mixing the APIs and excipients in a beaker and observed for 3,6 and 9 weeks. The mixture was checked for any physical or chemical changes in it.¹²⁻¹⁹

Calibration curve of drugs

A calibration curve for caffeine was prepared by dissolving 100 mg of caffeine in 100 mL of distilled water. Working solutions with varying concentrations were prepared by adding 0.1 mL of hydrogen chloride to each solution. The concentrations were determined using a UV spectrophotometer at 273 nm. To prepare a calibration curve for Ibuprofen, 20 mg of Ibuprofen were dissolving in 100 mL of phosphate buffer PBS, pH 6.8, using an ultra-sonifier. The stock solution was then used to prepare four different dilutions with concentrations of 0.05 mg/mL, 0.025 mg/mL, 0.0125 mg/mL and 0.0062 mg/mL. These dilutions were then analyzed using a UV-visible spectrophotometer at 222nm wavelength.²⁰

Method of Transdermal Patch Fabrication

The process of fabrication of transdermal patches was carried out through the solvent evaporation method. The formulation stage involved dissolving the matrix-forming polymer in 40 mL of distilled water through careful heating and vigorous stirring (Solution 1). Subsequently, a separate solution (Solution 2) was prepared by dissolving caffeine, SLS, Sodium benzoate and Tween-80 in 20 mL of distilled water, followed by the addition of Ibuprofen. Solution 2 was gradually introduced to Solution 1 with continuous stirring, resulting in the prepared solution being poured into the cast.

HPMC was prepared using warm distilled water to prevent gelation and polymers were prepared separately in 20 mL of distilled water. The drug concentration in the formulation influenced the release rate from the transdermal patch. The solution was spread into a Petri dish and the cast material was dried for four days to form a solid matrix. The dried material was then cut into 2x1 inch rectangular patches, affixed to a backing layer and packaged for stability. Assessments were conducted to verify drug content, patch thickness and adhesive properties, ensuring the patches were protected from environmental factors like moisture, heat and light. The formulation ingredients of all the nine formulations are listed along with their concentration in the following Table 1.

In vitro Characterization

Post-formulation studies

Transdermal patches were characterized for following parameters: physical appearance was examined visually, thickness was measured using micrometre screw gauge at three different points, weight uniformity was determined after drying at 60°C for 4 hr on a digital balance, content uniformity using spectrophotometry, flatness using by determination of % constriction, folding endurance by repeatedly folding a small strip of patch until it broke, % Moisture content using desiccator and weighing balance, surface pH was determined using pH meter, Drug content determination was done spectrophotometrically, shear adhesion test was conducted by hanging a specified weight with a steel plate to which patch was attached and peel adhesion test was done by pulling the patch at 180° after applying it to a steel plate, % elongation break test was determined by noting the length of the strip just before the breakpoint and Skin irritation was assessed using rabbit and Draiz scoring method for erythema.²¹⁻²⁴

Drug-Polymer Interaction

Excipients are an essential part of every pharmaceutical dosage form. The stability of formulation depends on various factors such as the compatibility of drugs with excipients. Both the drug and excipient must be compatible and stable with each other.^{16,25}

Fourier Transform Infra-Red spectroscopy (FTIR)

A FTIR spectrophotometer with an attenuated total reflectance accessory was used to analyze patches incorporated with drugs, pure crystalline drugs, polymers and physical mixtures. The powder sample was dried under vacuum to remove moisture effects and numerous scans were conducted at a resolution of 4 cm⁻¹ from 4000 cm⁻¹ to 600 cm⁻¹ for each spectrum.^{7,15}

Differential Scanning Calorimetry (DSC)

The DSC is a thermal analysis method for drugs using a liquid nitrogen cooling accessory. It uses Dry Nitrogen gas purging at 50 cc/min and high-purity indium for calibration. A 2.5-5 mg sample is prepared in an Aluminium crucible cell, heated to 250°C at 10°C per min and then analyzed.¹⁵

Thermogravimetric Analysis (TGA)

Thermogravimetric Analysis (TGA) is a quantitative method that measures the mass of a sample under furnace temperatures up to 1600°C. It detects mass reduction due to decomposition, volatile compound evaporation, or decreased oxidation state. When combined with Differential Scanning Calorimetry (DSC) or Differential Thermal Analysis (DTA), TGA helps deconvolute DSC plots by distinguishing physical changes from chemical ones.²⁶

X-ray Diffractometry studies (XRD)

XRD of medicated patches was taken. X-ray diffractometer containing 30 kV, 15 mA generator with Cu-K α anode tube. The diffraction pattern of medicated patches was scanned over the 2 θ range of 2° and 60° at 2°/min in 0.02° 2 θ step size.²⁷

Scanning Electron Microscopy

The SEM analysis involved using a Specify Model of SEM Instrument with specific details of equipment configuration. Transdermal patches were prepared, mounted on conductive stubs and coated with gold for better conductivity. SEM imaging was performed at different magnifications to capture detailed images of surface morphology and microstructure, with multiple areas examined for comprehensive representation.^{14,22}

Table 1: Nine formulations by varying different concentrations of ingredients.

Use	Ingredients	F1	F2	F3	F4	F5	F6	F7	F8	F9
Drugs	Caffeine	130 mg	130 mg	130 mg	130 mg	130 mg	130 mg	130 mg	130 mg	130 mg
	Ibuprofen	300 mg	300 mg	300 mg	300 mg	300 mg	300 mg	300 mg	300 mg	300 mg
Semi-Synthetic	HPMC	3 g	-	-	-	2 g	2 g	1g	1.5g	2 g
Natural Polymers	Tamarind Gum	-	8 g	-	4 g	-	4 g	3 g	3.5 g	4 g
	Moringa Gum	-	-	2 g	2 g	1 g	-	1 g	1.5 g	2 g
Solvent	Water	60 mL	60 mL	60 mL	60 mL	60 mL	60 mL	60 mL	60 mL	60 mL
Preservative	Na Benzoate	2 mg	2 mg	2 mg	2 mg	2 mg	2 mg	2 mg	2mg	2 mg
SAA+ Permeation Enhancer	SLS	100 mg	100 mg	100 mg	100 mg	100 mg	100 mg	100 mg	100 mg	100 mg
	Tween-80	2 mL	2 mL	2 mL	2 mL	2 mL	2 mL	2 mL	2 mL	2 mL

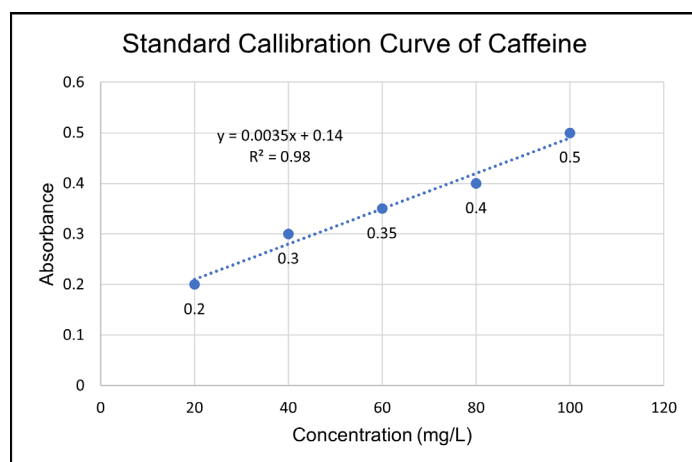


Figure 1: Standard calibration curve of caffeine.

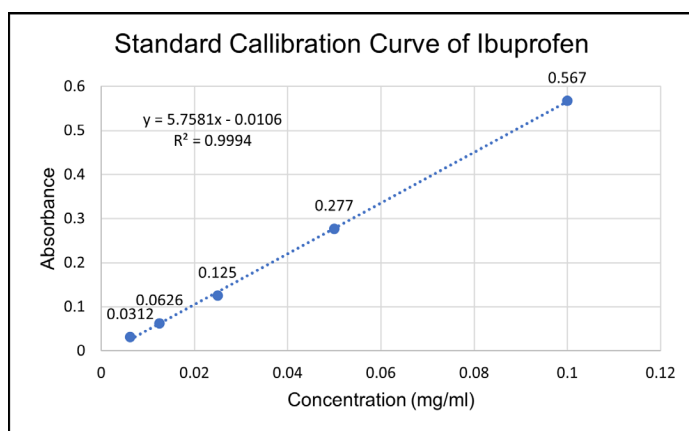


Figure 2: Standard calibration curve of Ibuprofen.

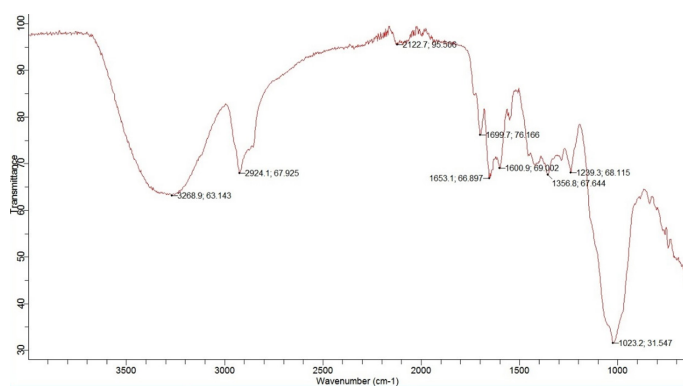


Figure 3: FTIR results of transdermal patches.

Drug Release Studies

Drug release studies are also essential for the characterization of the patch. In these studies, the Dissolution apparatus and Franz diffusion cell were used. These studies were conducted to check the *in vitro* release characteristics of the drug from the patch.^{10,24,25,28,29}

In vitro release studies

In vitro release studies can be done using a dissolution apparatus or Franz diffusion cell.

In vitro Dissolution

The dissolution study of a transdermal patch is conducted using a USP dissolution apparatus and a validated FDA Sandwich patch holder. The dissolution vessel is filled with 900 mL of distilled water, maintained at $37 \pm 0.5^\circ\text{C}$ and paddle speed at 50 rpm. The patch assembly is carefully placed and samples are withdrawn at 1 hr intervals until drug release is complete. The drug content is spectrophotometrically analyzed and plotted in a standard curve, revealing *in vitro* dissolution profiles, cumulative drug release and dissolution rate.^{3,6,19}

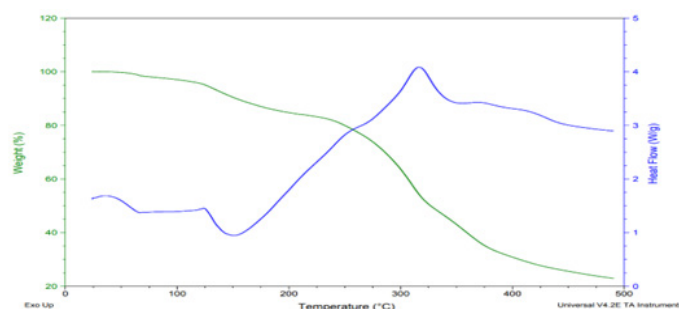


Figure 4: DSC and TGA thermogram of transdermal patches.

Release Kinetics

Various kinetic models can be used to determine the release kinetics, such as zero-order, first-order, Higuchi model and Korsmeyer-Peppas model. The coefficient of correlation (R^2) and rate constants can be determined using the Excel Add-In Program DD Solver1 and a comparison of R^2 values can be used to determine the best-fitted model.⁷⁻¹²

RESULTS AND DISCUSSION

Preformulation studies

Physicochemical characterization of drugs

The color of the caffeine was white; it was odorless and bitter in taste. The melting point of caffeine was 236.5°C ($234-239^\circ\text{C}$). Caffeine readily dissolved in water at room temperature and its solubility was observed to be 20 g/L at room temperature. The solubility in ethanol was 15 g/L at room temperature. The water/methylene chloride partition coefficient of caffeine was found to be 5.05. Ibuprofen was a colorless, crystalline and stable solid having a characteristic odor. The melting point was found to be 77°C . Ibuprofen exhibited insolubility in water but showed high solubility in acetone, methanol and methylene chloride. The value of the partition coefficient was found to be 3.97. Stability results showed that caffeine concentrations were within 5% of the expected concentration when stored for up to 6 months at room temperature.

Standard calibration curve of Caffeine

The standard dilutions were analysed at 273 nm through a UV Spectrophotometer. The standard calibration curve for caffeine is

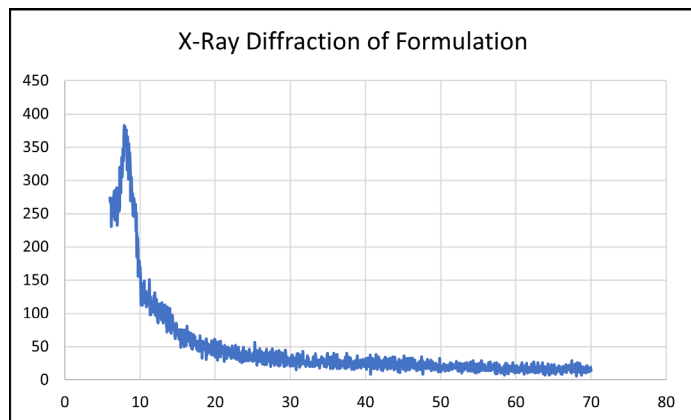


Figure 5: XRD diffractogram of transdermal patches.

presented in Figure 1.

Standard calibration curve of Ibuprofen

The standard dilutions were analysed at 222 nm. The standard calibration curve for ibuprofen is presented in Figure 2.

Post formulation studies

Evaluation of patches

The physical parameters of all patches are shown in Table 2, including weight, thickness, folding endurance, diameter, area, flatness and surface pH, were assessed. The patches, sized 2×1 inches, demonstrated flexibility, uniformity, smoothness and transparency. The weight ranged from 2.049 to 2.224 g, while the thickness varied from 0.2 to 0.5 mm. Folding endurance exceeded 300, indicating strong elasticity. Flatness study confirmed 100% flatness and moisture absorption rates were minimal (0.0012 to 0.0055%), ensuring stability and microbial

Table 2: Physiochemical Characterization of Patches.

Formulation code	Physical appearance	Thickness (mm)	Weight (g)	Flatness (%)	Folding endurance	% Moisture content	pH	Drug content (%)	Tensile strength (kg/cm ²)	Elongation at break (%mm ⁻²)
F1	++	0.204	2.049	100	>200	0.0034	7.0	96.64 ± 0.5	0.31 ± 0.038	107.34 ± 19.346
F2	++	0.312	2.145	100	>200	0.0052	7.0	97.68 ± 0.2	1.47 ± 0.051	18.48 ± 3.490
F2	++	0.224	2.057	100	>200	0.0024	7.0	98.45 ± 0.2	0.83 ± 0.047	31.70 ± 3.611
F4	++	0.514	2.214	100	>200	0.0014	7.0	97.94 ± 0.3	0.66 ± 0.208	64.78 ± 11.072
F5	++	0.437	2.050	100	>200	0.0012	7.0	98.79 ± 0.3	0.56 ± 0.014	73.34 ± 10.828
F6	++	0.226	2.198	100	>200	0.0046	7.0	98.45 ± 0.2	2.20 ± 0.091	11.53 ± 1.876
F7	++	0.585	2.221	100	>200	0.0038	7.0	99.73 ± 0.4	2.15 ± 0.077	12.02 ± 0.862
F8	++	0.234	2.224	100	>200	0.0054	7.0	98.46 ± 0.3	1.58 ± 0.052	15.26 ± 1.231
F9	++	0.322	2.071	100	>200	0.0055	7.0	99.61 ± 0.5	1.31 ± 0.311	26.75 ± 8.371

++ = Satisfactory results.

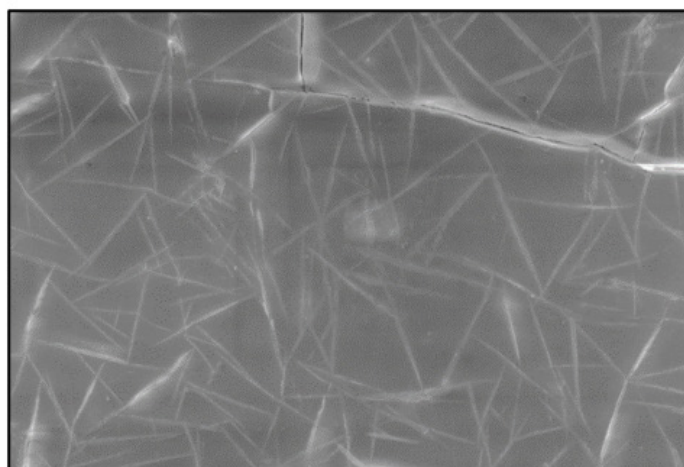
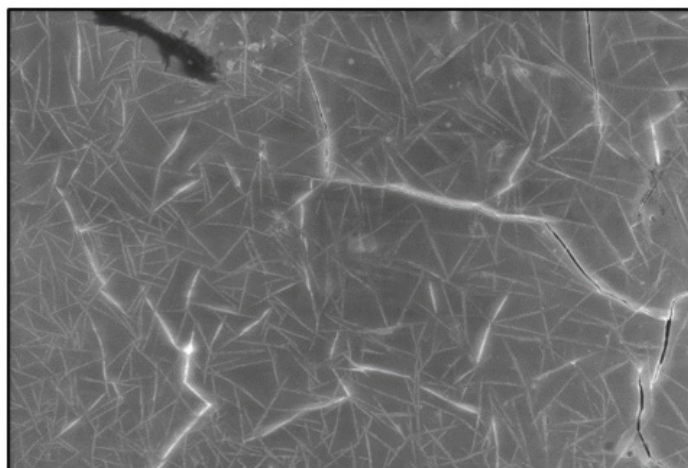


Figure 6: SEM results at 500x and 1kx of transdermal patches matrix.

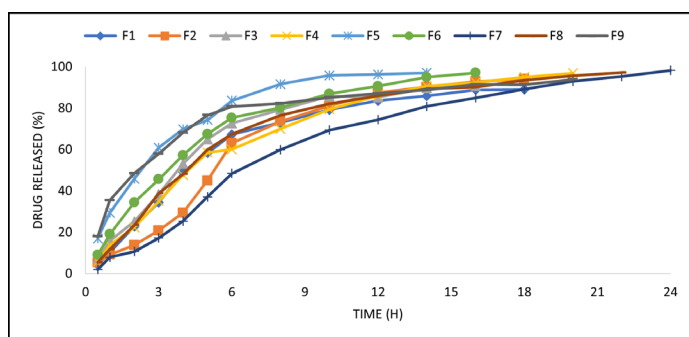


Figure 7: % Drug release profile of transdermal patches.

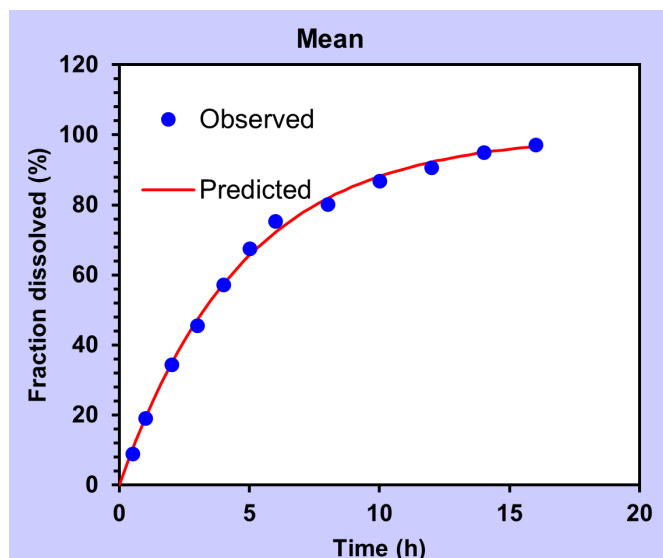


Figure 9: First order.

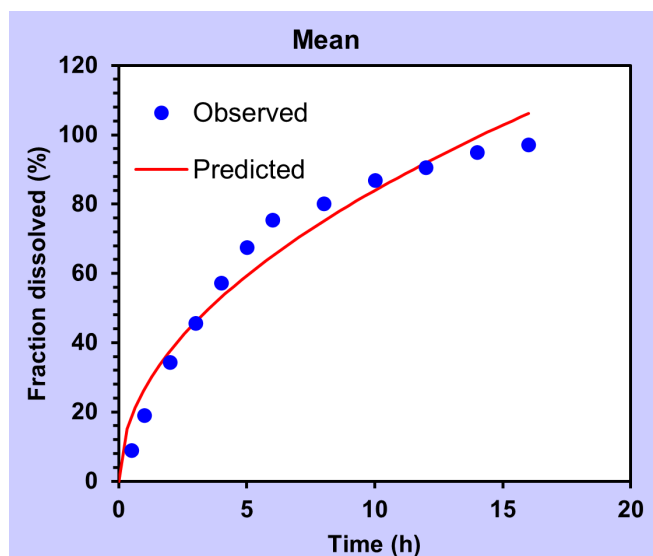


Figure 10: Higuchi.

protection. Moisture loss ranged from 1.84 to 7.34%. The surface pH was 5.5 ± 0.08 , aligning with skin pH to prevent irritation. Drug content uniformity ranged from 96.28 to 99.73%, indicating a consistent manufacturing process. Shear strength for patches

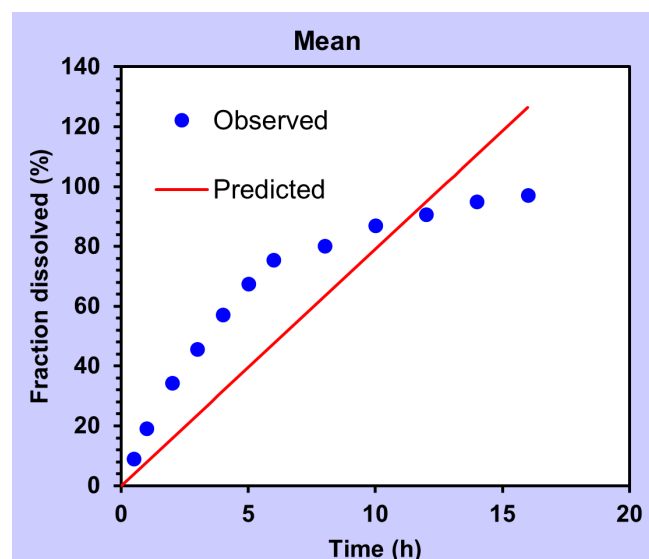


Figure 8: Zero order.

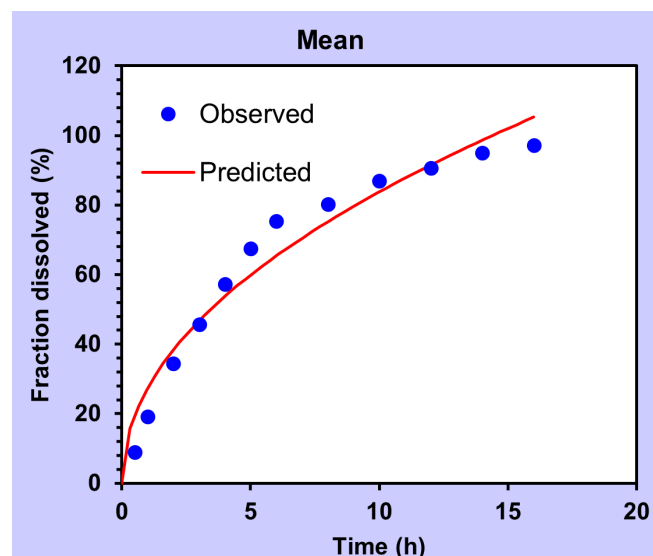


Figure 11: Korsmeyer-Peppas.

with permeation enhancers was 20-30 min and peel strength demonstrated strong anchorage and cohesion. % Elongation ranged from $28.95 \pm 0.015\%$ to $41.2 \pm 0.015\%$. Skin irritancy study on rabbits showed no erythema or oedema, confirming skin compatibility.

Drug polymer interactions

FTIR of Formulation

The FTIR results of the formulation, as shown in Figure 3, showed that it was made up of all three polymers namely Moringa, Tamarind and HPMC as at 3268.9 cm^{-1} due to stretching vibration of -OH group, 2924 cm^{-1} due to C-H stretching of arabinose and 1600.9 cm^{-1} due to C=O stretching which is indicative of the features of Moringa and tamarind gum. The peak at 1356.8 cm^{-1} is due to the bending vibration of the -OH group and 1023

cm⁻¹ is due to C=O stretching vibration, showing the presence of HPMC in the formulation matrix. There were no characteristic peaks of drugs (Caffeine and Ibuprofen) seen in the results of the formulation which can be inferred that the drug is suitably incorporated in the matrix system of these polymers.

Differential Scanning Calorimetry (DSC)

The differential scanning calorimetry thermal analysis of the patch is presented in Figure 4, which shows no change in temperature up to 100°C, but after that, it shows a downward step curve, which indicates an endothermic trough at 150°C. It shows the

crystallinity and purity of ibuprofen. This observation confirms the presence of ibuprofen's physical and chemical characteristics. After that, the upward curve at 325°C indicates exothermic change, which means heat was released caused by crystallization.

Thermogravimetric Analysis (TGA)

The gravimetric analysis of the patch, presented in Figure 4, showed no change in weight loss up to 100°C, which indicated the patch's thermal stability. After 100°C, the curve stepped down to 250°C, which showed that degradation had started. After 250°C, there was a sharp decline, showing the patch's degradation. Complete degradation was observed at 500°C.

Table 3: % Drug release profile of transdermal patches.

Time (Hr)	% Drug Release Profile								
	F1	F2	F3	F4	F5	F6	F7	F8	F9
0.5	4.11	5.4	7.0	5.9	17.0	9.0	5.4	2.0	17.9
1	10.3	9.2	15.9	13.8	29.4	19.1	11.8	7.9	35.4
2	22.8	13.8	25.2	22.3	45.8	34.4	23.4	10.7	48.5
3	34.4	20.8	38.4	35.0	60.9	45.6	38.9	17.2	57.9
4	49.1	29.4	53.1	47.7	69.8	57.2	48.1	25.4	68.1
5	58.3	45.0	65.0	58.5	74.3	67.5	60.0	37.1	76.6
6	67.3	63.1	72.7	60.1	83.7	75.4	67.5	48.4	80.8
8	73.0	73.6	79.3	70.0	91.6	80.2	76.5	59.9	82.3
10	79.2	81.1	85.8	79.5	95.8	86.9	82.1	69.4	85.2
12	83.6	87.5	85.3	86.0	96.3	90.6	86.0	74.4	87.1
14	85.9	90.3	89.8	90.6	97.1	95.0	89.4	80.9	88.9
16	88.9	92.9	90.8	92.4		97.1	90.2	85.0	91.5
18	89.0	94.1		95.1			93.5	89.1	91.4
20				96.9			95.7	93	93.9
22							97.2	95.3	
24								98.3	

Table 4: Release Kinetics of transdermal patches.

Formulation	Zero Order		First Order		Higuchi		Korsmeyer	
	K ₀	R ²	K1	R ²	kH	R ²	kKP	n
F1	6.517	0.6375	0.157	0.9852	23.111	0.9272	21.699	0.412
F2	6.567	0.6375	0.138	0.9515	22.703	0.8892	14.808	0.421
F3	7.500	0.5978	0.185	0.9849	25.067	0.9246	24.179	0.321
F4	6.217	0.6795	0.157	0.9954	23.232	0.9574	21.565	0.422
F5	9.527	0.1626	0.304	0.9951	30.250	0.9122	36.437	0.409
F6	7.905	0.5323	0.213	0.9975	26.533	0.9510	27.400	0.443
F7	5.813	0.5299	0.168	0.9932	23.050	0.9296	24.488	0.419
F8	4.985	0.8668	0.108	0.9755	20.051	0.9248	13.009	0.398
F9	6.502	-0.7843	0.280	0.9124	25.377	0.6624	41.235	0.298

X-ray Diffractometry Study

Diffraction measurements were conducted using a high-resolution diffractometer; resulting diffractograms illustrated the transition from b to a-caffeine between 130 and 140°C, with characteristic peaks disappearing and emerging. In a separate study involving Ibuprofen, all crystal forms were subjected to powder X-ray diffraction pattern analysis using a Rigako model DIMAX-RC diffractometer. Alterations in peak locations, appearances and disappearances were observed, indicating changes in crystal forms, as shown in Figure 5. Distinct polymorphic forms were evidenced by variations in interplanar distance (d values) and diffraction relative Intensities (I/I₀). The preparation of a transdermal patch containing caffeine and ibuprofen involved the dissolution of both Active Pharmaceutical Ingredients (APIs) in polymers such as moringa gum, tamarind gum and HPMC. Formulated through the solvent evaporation method, XRD analysis demonstrated the complete incorporation of APIs into the polymer matrix, with no distinct sharp peaks observed, indicative of the successful integration of drugs in the matrix.

Scanning Electron Microscopy

Microscopy studies are helpful for observing the surface morphology of the matrix generated and evaluating the physical stability of the formulations. SEM studies were conducted to visualize drug distribution in the matrix patch. The SEM studies clearly showed that a reasonable matrix was formed with a uniform repeating three-dimensional mesh in which the drug is evenly distributed. The SEM results are shown in Figure 6.

Drug release studies

In vitro Dissolution

The dissolution test was performed using the Dissolution apparatus USP under suitable conditions at 37°C for a specified time. Drug release characteristics of formulations were studied in *in vitro* conditions. Formulations F1, F2 and F3 have shown drug release of about 4.11%, 5.4% and 7.0% respectively in the first 30 min. The maximum drug released from formulations F1, F2 and F3 was 89.0%, 94.1% and 90.8%, respectively, with a duration of 18, 18 and 16 hr, respectively. The formulations F4, F5 and F6 showed drug release of 5.9%, 17.0% and 9.0%, respectively, in the first 30 min. The overall duration for drug release from formulations F4, F5 and F6 was found to be 20, 14 and 16 hr, respectively. The maximum drug release from formulations F4, F5, F6 is 96.9%, 97.1%, 97.1% respectively. Formulations F7, F8 and F9 showed a drug release of about 5.4%, 2.0% and 17.9% respectively. The drug release duration from formulations F7, F8 and F9 was found to be 22, 24 and 20 hr, respectively. The maximum drug release from formulations F7, F8, F9 is 97.2%, 98.3%, 93.9% respectively. The order of drug release was F8>F7>F4>F9>F2>F1> F3> F5.

The best formulation was found to be F7 and F8 having varying concentrations of *Moringa* gum, tamarind gum and HPMC which gave the desired drug release for 24 and 22 hr respectively because polymers were used in combination which exhibited this sustained behavior. All the formulations followed the first-order drug release. The % Drug release profile of transdermal patches is shown in Figure 7 and Table 3.

In vitro drug release kinetics

The *in vitro* permeation profile data of various formulations through Franz diffusion cells was subjected to analysis for different release kinetics, including zero-order, first-order, Higuchi-type and Korsmeyer-Peppas-type release kinetics. The results of all formulations are presented in Table 4. The coefficient of correlation (R^2) was calculated for each release kinetics model and a comparative assessment was conducted. Notably, the first-order kinetics demonstrated higher R^2 values (ranging from 0.9124 to 0.9975) compared to zero order (ranging from -0.7843 to 0.8668) and Higuchi's square-root model (ranging from 0.6624 to 0.9674). This observation suggested a concentration-dependent drug release rate from the formulations, with formulation F6 exhibiting the highest R^2 value of 0.9975, signifying its superior performance.

The application of the Korsmeyer-Peppas model to the release profiles facilitated the determination of the diffusional exponent (n), indicative of the release mechanism. Notably, all formulations exhibited 'n' values of 0.412, 0.4210, 0.321, 0.422, 0.409, 0.443, 0.419, 0.398 and 0.298 for F1, F2, F3, F4, F5, F6, F7, F8 and F9, respectively consistent with Fickian diffusion. A 'n' value of 0.45 or below suggested Fickian diffusion, while values between 0.45 and 1.00 indicate non-Fickian transport and values exceeding 1.00 suggest zero order/case II transport. The obtained 'n' values support the conclusion that the drug releases conform to Fick's law of diffusion across all formulations. The graphical presentation of Release kinetics of formulation is shown in Figures 8-11 below.

CONCLUSION

The transdermal patch for the prolonged management of migraine using natural polymers *Moringa*, Tamarind gum and a semi-synthetic polymer HPMC. Nine formulations were prepared and all of them showed satisfactory dissolution and release kinetic results and followed Fickian diffusion. A successful matrix system was fabricated and confirmed with Scanning Electron Microscope (SEM) results and the drug was incorporated in the three-dimensional mesh. It was concluded that transdermal patches showed a sustained drug release pattern over 24 hr. We can modulate the duration of the sustained drug release exceeding 24 hr by simply increasing the concentration of matrix-forming polymers, which will retard the drug release to the desired level.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ABBREVIATIONS

HPMC: Hydroxypropyl Methylcellulose, **TDD:** transdermal drug delivery, **TDDS:** transdermal drug delivery system, **COX:** Cyclooxygenase enzyme, **CMC:** Carboxy methyl cellulose, **SLS:** Sodium lauryl sulphate, **MP:** Melting point, **PBS:** Phosphate buffer solution, **SAA:** Surface active agent, **FDA:** Food and drug administration, **SEM:** Scanning electron microscope.

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